

HUMAN INTESTINAL DENDRITIC CELLS DICTATE INFLAMMATION AND T-CELL POLARIZATION

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Introduction: Enormous diversity of commensal bacteria determines individual functions acting on the development and activities of the human immune system. This complexity can directly be translated to T-cell polarization to support tolerance induction or inflammation. We have established a sensitive *in vitro* culture system for investigating the response of monocyte-derived dendritic cell (moDC) subsets to various gut bacteria by monitoring the expression of type I/II CD1 proteins, secretion of chemokines, pro-inflammatory and T-cell polarizing cytokines in the context of the T-cell polarizing potential of these moDC subsets. Under physiological conditions the gut microenvironment is conditioned by all-transretinoic acid (ATRA) produced by gut epithelial cells and CD103⁺ DCs. To consider the impact of this special microenvironment on moDC-induced T-cell responses we compared the effects of selected microbes on DC and T-cell development in the absence and presence of ATRA.

Methods: Monocytes were separated from human buffy coats and differentiated *in vitro* in the presence of IL-4 and GM-CSF with or without 1 nM ATRA for 2 days. Gram(-) (*Schaedler's E. coli*, *E. coli* 058, *M. morganii*) and Gram(+) (*B. subtilis*) bacteria were grown in antibiotic-free LB medium and were added to the 2-day moDCs for 24 hrs. Activation of moDC was monitored by the expression of membrane CD1a, CD1d and CD83 by FACS analysis. Culture supernatants of activated moDC were collected on day 3 and cytokine concentrations were determined by ELISA. The number of IFN γ and IL-17 producing T-cells was measured by ELISPOT assay. Expression levels of selected NOD-like receptors and genes involved in ATRA synthesis were measured by qRT-PCR.

Results: Increased expression of CD83 revealed that all tested commensal bacteria were able to activate moDCs for pro- and anti-inflammatory cytokine secretion. ATRA had a significant impact on the differentiation, inflammatory response and T-cell polarizing activity of moDCs. It increased the cell surface expression of CD1a while increased that of CD1d, previously shown to be associated with a shift in moDC functionality. ATRA also enhanced IL-1 β secretion and upregulated the expression of genes involved in ATRA synthesis and NLRP12 mRNA levels. Interestingly, these ATRA-induced effects could be counterregulated by the tested microbe. The interaction of microbes resulted in IL-23 production supporting Th17 polarization of autologous T-cells and increased the number of IFN γ producing T-cells however, these effects were down modulated by ATRA.

Discussion: In our culture system we identified two moDC subpopulations referred as DC-SIGN⁺CD11c⁺CD14^{med}CD1a⁺CD1d⁻ and DC-SIGN⁺CD11c⁺CD14⁺CD1a⁻CD1d⁺ cells, which respond to and coordinate stimuli from commensal bacteria differently to induce T-cell polarization and expansion. Our results also showed that the tested bacteria modulate the differentiation and activation of moDCs in a dose- and bacterial strain-dependent manner. Moreover, the interplay of moDC and commensals can be modified by the actual milieu of the cells such as ATRA.